

Structural model for the Cu_B site of dopamine β-hydroxylase and peptidylglycine α-hydroxylating monooxygenase: crystal structure of a copper(II) complex showing N₃OS coordination and axial sulfur ligation

Bidyut K. Santra, Pattubala A. N. Reddy, Munirathinam Nethaji and Akhil R. Chakravarty*

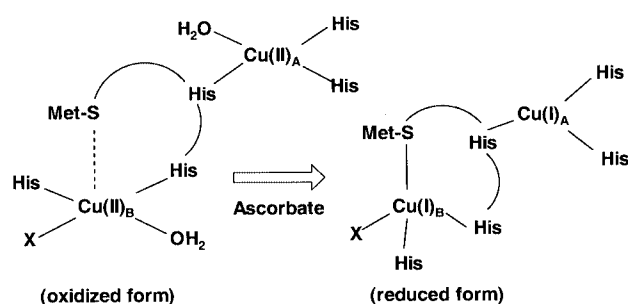
Department of Inorganic & Physical Chemistry, Indian Institute of Science, Bangalore -560012, India. E-mail: arc@ipc.iisc.ernet.in

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A copper(II) complex [Cu(L)(phen)](ClO₄) (HL, NSO-donor Schiff base ligand) with a Cu^{II}N₃OS geometry showing axial sulfur ligation is a structural model for the Cu_B site of DβH and PHM, and the complex is catalytically active in the oxidation of ascorbic acid by dioxygen mediated by a copper(I) species.

Dopamine β-hydroxylase (DβH; E.C.1.14.17.1) and peptidylglycine α-hydroxylating monooxygenase (PHM; E.C.1.14.17.3) are copper proteins responsible for the benzylic hydroxylation of dopamine to norepinephrine in the biosynthesis of the neurohormone adrenaline and α-amidating bioactive peptides, respectively.^{1–3} The active sites of DβH and PHM have structural and functional similarities. The enzymes have two essentially uncoupled copper centres per subunit. The crystal structure of PHM shows a separation of 11 Å between two mononuclear copper sites.⁴ While the Cu_A site mediates electron transfer, the Cu_B site is responsible for the functionalization of the organic substrate. The Blackburn model for the oxidized and reduced active sites of DβH and PHM shows a CuN₃OS coordination for the Cu(II)_B site with the MetS ligand occupying the axial site (Scheme 1).⁵ The axially bound Met residue



Scheme 1 Blackburn model showing the active site coordination geometries in the oxidized and reduced forms of DβH and PHM [Cu(II)_BN₃OS geometry with X as either His or an azide ion].

is known to be crucial for the catalytic activity.⁶ The crystal structure of PHM shows the presence of a weakly bound Met residue at a distance of 2.68 Å.⁴ Herein, we present the synthesis, crystal structure and properties of a copper(II) complex, [Cu^{II}(L)(phen)](ClO₄) (**1**) which shows axial sulfur ligation in a CuN₃OS geometry giving a similar Cu–S distance as in PHM. Complex **1** is of significance as a structural model for the Cu(II)_B site of DβH and PHM as the presently known⁷ copper(II) complexes with N₃OS coordination have sulfur ligation at the basal plane.

The copper(II) complex [Cu(O-2-C₆H₄CH=NC₆H₄-2'-SMe)-(phen)](ClO₄) (**1**) was prepared from the reaction of copper(II) acetate hydrate with 1,10-phenanthroline (phen) and 2-(methylthiophenyl)salicylaldehyde (HL) in methanol followed by addi-

tion of sodium perchlorate. The copper(I) complex [Cu(HL)-(phen)](ClO₄) (**2**) was obtained by reducing **1** with ascorbic acid (H₂A) in an aqueous acetonitrile medium under anaerobic conditions.[†] Both the complexes show an infrared band near 1090 cm^{–1} assignable to the perchlorate anion. Complex **1** displays a d–d band at 646 nm and a charge transfer transition at 402 nm in MeCN. Complex **2** exhibits a charge transfer transition at 348 nm with a shoulder at 536 nm in MeCN. Complex **1** is one-electron paramagnetic and shows an axial EPR spectrum with $g_{\parallel} = 2.16$ ($A_{\parallel} = 114 \times 10^{-4}$ cm^{–1}) and $g_{\perp} = 2.02$ in MeCN–toluene glass (1 : 1 v/v). Complex **2** is diamagnetic and exhibits a singlet at 13.19 ppm in the ¹H NMR spectrum in CD₃CN assignable to the phenolic OH. The non-involvement of the phenolic group in **2** in metal binding is evidenced from the negligible shift of the proton resonance compared to that of the free ligand [OH(ligand) = 13.16 ppm]. The formation of a copper–sulfur bond in **2** is indicated from the significant downfield shift of the S-methyl resonance from 2.28 ppm in HL to 2.52 ppm in **2**.^{8,9} The ¹H NMR data suggest a four-coordinate geometry for **2** with a CuN₃S coordination geometry which models the Cu_B site structure of DβH and PHM in the reduced state.

The crystal structure of **1** shows a tridentate NSO coordination of the Schiff base and a bidentate NN mode of bonding for the phen ligand (Fig. 1).[‡] The coordination geometry is

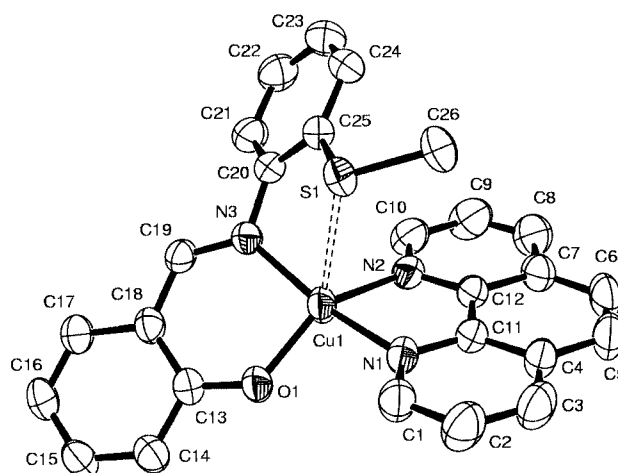


Fig. 1 An ORTEP view of the cation of complex **1** with thermal ellipsoids at the 50% probability level and the atom numbering scheme. Selected bond distances (Å) and angles (°): Cu(1)–N(1) 1.995(3), Cu(1)–N(2) 2.069(3), Cu(1)–N(3) 1.949(3), Cu(1)–O(1) 1.877(3), Cu(1)–S(1) 2.765(1); O(1)–Cu(1)–N(1) 91.66(12), O(1)–Cu(1)–N(2) 142.77(12), O(1)–Cu(1)–N(3) 95.25(11), O(1)–Cu(1)–S(1) 128.65(9), N(1)–Cu(1)–N(2) 81.52(12), N(1)–Cu(1)–N(3) 162.28(13), N(1)–Cu(1)–S(1) 89.01(10), N(2)–Cu(1)–N(3) 102.23(12), N(2)–Cu(1)–S(1) 88.01(9), N(3)–Cu(1)–S(1) 73.94(9).

distorted square-pyramidal with three nitrogen and one oxygen atoms occupying the basal plane ($\tau = 0.3$). The Cu(1)–O(1) bond length of 1.877(3) Å is the shortest in the basal plane. The sulfur atom is involved in a weak axial ligation giving a Cu–S distance of 2.765(1) Å. The bond length compares well with the Cu_B–M314 distance of 2.68 Å found in the crystal structure of PHM.⁴ Complex **1** with axial sulfur coordination in a CuN₃OS geometry exemplifies the first structural model for the Cu_B site of DβH and PHM in the oxidized form. A recent report on the modeling of the Cu_B site shows axial sulfur ligation in a copper(II) complex having a CuN₃SCl coordination geometry.¹⁰

The redox activity of the complexes has been studied by cyclic voltammetry in both protic and aprotic solvents. In dmf–Tris–HCl/0.1 M KCl buffer medium [1 : 4 v/v, pH 7.2, Tris = tris(hydroxymethyl)aminomethane], the complexes show a quasi-reversible voltammetric response for the Cu(II)/Cu(I) couple at –0.10 and –0.08 V for **1** and **2** respectively, with a ΔE_p value of 120 mV at a 50 mV s^{–1} scan rate (Fig. 2). The ratio

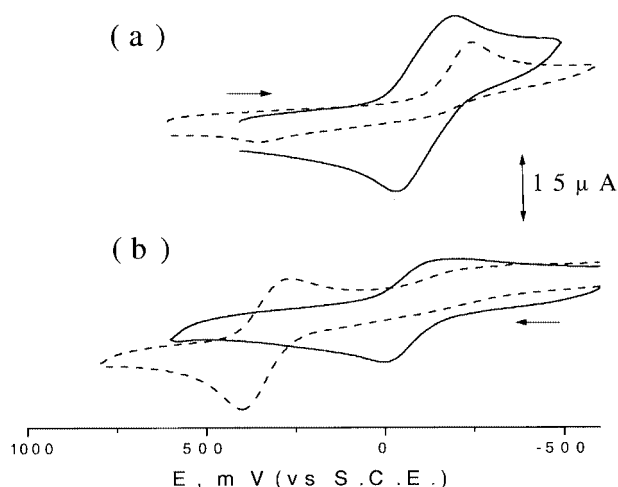
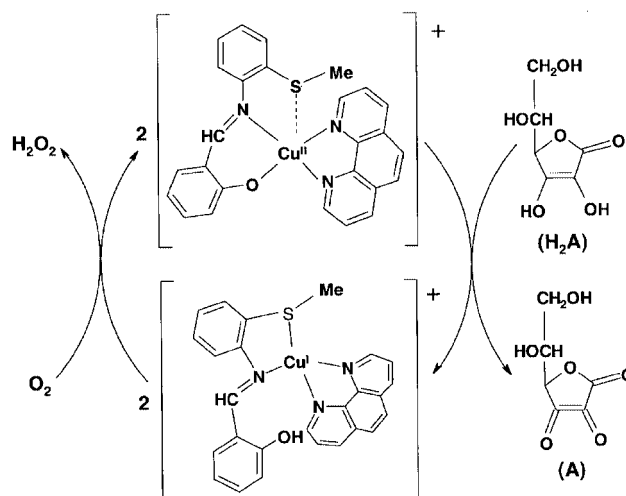


Fig. 2 Cyclic voltammograms of [Cu^{II}(L)(phen)](ClO₄) (**1**, a) and [Cu^I(HL)(phen)](ClO₄) (**2**, b) in dmf–Tris–HCl/0.1 M KCl buffer (1 : 4 v/v, pH 7.2) (—) and CH₂Cl₂–0.1 M TBAP (---) at a scan rate of 50 mV s^{–1}.

of anodic (i_{pa}) and cathodic (i_{pc}) peak currents is unity at scan rates of 50–200 mV s^{–1}. In an aprotic solvent such as CH₂Cl₂–0.1 M TBAP (tetrabutylammonium perchlorate), complex **1** displays an irreversible reduction peak at –0.25 V due to the formation of [Cu^I(L)(phen)] (1[–]) (Fig. 2a). An anodic response with much reduced current ($i_{pa}/i_{pc} \approx 0.1$) is observed at 0.35 V. This anodic peak is assignable to the oxidation of complex **2** formed in trace quantity from the unstable 1[–] species during the voltammetric scan. A better reversibility for the Cu(II)/Cu(I) couple in the buffer medium could be due to the presence of an electroprotic reaction: [Cu^{II}(L)(phen)]⁺ + H⁺ + e[–] ⇌ [Cu^I(HL)(phen)]⁺. In CH₂Cl₂–0.1 M TBAP complex **2** exhibits a quasireversible voltammogram at 0.34 V ($\Delta E_p = 120$ mV) with a i_{pa}/i_{pc} ratio of unity due to the Cu(II)/Cu(I) couple involving [Cu^{II}(HL)(phen)]²⁺ and [Cu^I(HL)(phen)]⁺ (Fig. 2b). An additional cathodic response near –0.25 V with a reduced peak current is observed, possibly due to the formation of a trace quantity of **1** during oxidation of complex **2**. The high positive potential for the Cu(II)/Cu(I) couple in **2** in CH₂Cl₂ is due to stabilization of the copper(I) state in a CuN₃S coordination geometry.

Complex **1** readily reacts with ascorbic acid (H₂A) to form the reduced complex **2**. The reduced species converts to **1** on exposure to dioxygen. This cyclic process is effective with a 1 : 100 mol ratio of **1** and H₂A in a dmf–Tris–HCl buffer (1 : 4 v/v, pH 7.2). In the presence of excess H₂A, **1** degrades to a non-catalytic copper(II) species. The catalytic process is of significance as it involves two copper species having Cu^{II}N₃OS and Cu^IN₃S coordination geometries that model the structural changes associated with the Cu_B site of DβH and PHM in enzymatic reactions (Scheme 2).



Scheme 2 Catalytic oxidation of ascorbic acid by dioxygen mediated by five-coordinate [Cu^{II}(L)(phen)](ClO₄) and four-coordinate [Cu^I(HL)(phen)](ClO₄) species in a dmf–Tris–HCl buffer (1 : 4 v/v, pH 7.2).

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Notes and references

† The complex [Cu(L)(phen)](ClO₄) (**1**) was prepared in 75% yield from the reaction of Cu₂(O₂CMe)₄(H₂O)₂ (0.5 mmol) with phen (1.0 mmol) in 15 cm³ MeOH. The mixture was left stirring for 0.5 h at 25 °C at which point Schiff base (HL, 1.0 mmol), obtained from the condensation of salicylaldehyde and 2-(methylthio)aniline, was added. The complex was isolated as a green solid by addition of a methanolic solution of NaClO₄ (1.0 mmol). Found: C, 53.6; H, 3.6; N, 7.5. Calc. for C₂₆H₂₀N₃O₅SClCu (**1**): C, 53.3; H, 3.4; N, 7.2%. λ_{max}/nm (ε/dm³ mol^{–1} cm^{–1}) in MeCN: 646 (190), 402 (6350), 270 (42900), 225 (51200). Complex **2** was isolated in 70% yield as a brown solid by reacting complex **1** (0.34 mmol) in MeCN (15 cm³) with ascorbic acid (0.34 mmol) in H₂O (1 cm³) under a dinitrogen atmosphere. Solvents were removed under vacuum and the solid was thoroughly washed with deoxygenated cold water and finally dried *in vacuo* over P₄O₁₀. Found: C, 53.6; H, 4.0; N, 7.5. Calc. for C₂₆H₂₁N₃O₅SClCu (**2**): C, 53.2; H, 3.6; N, 7.2%. λ_{max}/nm (ε/dm³ mol^{–1} cm^{–1}) in MeCN: 536 sh, 348 (11400), 267 (52200), 225 (61600). ¹H NMR in CD₃CN, δ: 2.52 (s, 3H, S–Me), 7.01–7.57 (16H, aromatic protons), 8.84 (s, N=CH), 13.19 (s, OH) (s, singlet). **CAUTION!** Perchlorate salts are potentially explosive. Single crystals of **1**, suitable for X-ray studies, were obtained by slow evaporation of an aqueous methanolic solution of the complex.

‡ Crystal data for **1**: C₂₆H₂₀N₃O₅SClCu, *M* = 585.50, triclinic, space group *P* $\bar{1}$ (no. 2), *a* = 9.366(2), *b* = 10.706(6), *c* = 13.686(3) Å, *a* = 100.20(3), *β* = 105.57(2), *γ* = 101.83(3)°, *U* = 1254.1(8) Å³, *Z* = 2, *D_c* = 1.551 g cm^{–3}, *T* = 293(2) K, 1.59 ≤ *θ* ≤ 24.97°, *μ* = 11.04 cm^{–1}, *R*(000) = 598, *R*1 = 0.0461, *wR*2 = 0.1514 for 3929 reflections with *I* > 2σ(*I*) and 415 parameters [*R*1 (*F*²) = 0.0506 (all data)]. Weighting scheme: *w* = 1/[σ²(*F_o*²) + (0.0969*P*)² + 1.0991*P*], where *P* = (*F_o*² + 2*F_c*²)/3. Intensity data from a crystal of dimensions 0.41 × 0.36 × 0.16 mm were obtained on an Enraf-Nonius CAD4 diffractometer using graphite-monochromated Mo–Kα radiation (λ = 0.7107 Å). Data were corrected for Lorentz, polarization and absorption effects. Structure solution and refinement were performed using the SHELX system of programs.¹¹ The perspective view of the molecule was obtained using ORTEP.¹² CCDC reference number 172492. See <http://www.rsc.org/suppdata/dt/b1/b109332k/> for crystallographic data in CIF or other electronic format.

- J. P. Klinman, *Chem. Rev.*, 1996, **96**, 2541; A. G. Blackman and W. B. Tolman, *Struct. Bonding (Berlin)*, 2000, **97**, 179.
- L. C. Stewart and J. P. Klinman, *Annu. Rev. Biochem.*, 1988, **57**, 551.
- J. S. Boswell, B. J. Reedy, R. Kulathila, D. Merkler and N. J. Blackburn, *Biochemistry*, 1996, **35**, 12241.
- S. T. Prigge, A. S. Kolhekar, B. A. Eipper, R. E. Mains and L. M. Amzel, *Science*, 1997, **278**, 1300.

- 5 B. J. Reedy and N. J. Blackburn, *J. Am. Chem. Soc.*, 1994, **116**, 1924.
- 6 B. A. Eipper, A. S. W. Quon, R. E. Mains, J. S. Boswell and N. J. Blackburn, *Biochemistry*, 1995, **34**, 2857.
- 7 E. W. Arinscough, E. N. Baker, A. M. Brodie, R. J. Cresswell, J. D. Ranford and J. M. Waters, *Inorg. Chim. Acta*, 1990, **172**, 185; L. Latos-Grazynski, J. Lisowski, M. M. Olmsted and A. L. Balch, *J. Am. Chem. Soc.*, 1987, **109**, 4428; S. Gou, X. You, Z. Xu, Z. Zhou and K. Yu, *Polyhedron*, 1991, **10**, 1363; H. E. Heldal and J. Sletten, *Acta Chem. Scand.*, 1996, **50**, 596; M. Ruf and C. G. Pierpont, *Angew. Chem., Int. Ed.*, 1998, **37**, 1736; M. M. Whittaker, Y.-Y. Chuang and J. M. Whittaker, *J. Am. Chem. Soc.*, 1993, **115**, 10029; I. Castro, M. L. Calatayud, J. Sletten, F. Lloret, J. Caro, M. Julve, G. Seitz and K. Mann, *Inorg. Chem.*, 1999, **38**, 4680.
- 8 K. D. Karlin, M. S. Haka, R. W. Cruse, G. J. Meyer, A. Farooq, Y. Gultneh, M. S. Hayes and J. Zubietta, *J. Am. Chem. Soc.*, 1988, **110**, 1196.
- 9 F. Champloy, N. Benali-Chérif, P. Bruno, I. Blain, M. Pierrot and M. Réglér, *Inorg. Chem.*, 1998, **37**, 3910; R. R. Gagne, R. P. Kreh, J. A. Dodge, R. E. March and M. McCool, *Inorg. Chem.*, 1982, **21**, 254; S. M. Nelson, A. Lavery and M. G. B. Drew, *J. Chem. Soc., Dalton Trans.*, 1986, 911.
- 10 M. Kodera, T. Kita, I. Miura, N. Nakayama, T. Kawata, K. Karo and S. Hirota, *J. Am. Chem. Soc.*, 2001, **123**, 7715.
- 11 G. M. Sheldrick, SHELX-97, Programs for Crystal Structure Solution and Refinement, University of Göttingen, Germany, 1997.
- 12 C. K. Johnson, ORTEP, Report ORNL-5138, Oak Ridge National Laboratory, Oak Ridge, TN, 1976.